

## Response of common bean (*Phaseolus vulgaris* L.) plant to foliar spray with some growth activators

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### Abstract

Two field experiments were conducted at the experimental Farm Station of Hort. Faculty of Agriculture Moshtohor, Benha University, Qalubia Governorate, Egypt, during the seasons of 2022 and 2023 This study aimed to investigate the effect of foliar spray with some growth activators i.e., benzyladenine at 25 and 50 mg L<sup>-1</sup>, milagrow at 50 and 100 mg L<sup>-1</sup>, yeast extract at 50 and 100 ml L<sup>-1</sup> as well as lithovit at 250 and 500 mg L<sup>-1</sup> on growth, chemical composition, anatomical features as well as yield characteristics of common bean plant.

The obtained results indicated that, significant increases existed in many growth characteristics and bioconstituents level such as photosynthetic pigments, phytohormones, minerals, total carbohydrates as well as crude protein %. Also, results showed that applied treatments enhanced anatomical features of common bean plant. Moreover yeast extract at 100 ml L<sup>-1</sup> and milagrow at 100 mg L<sup>-1</sup> treatments caused significant increases in flowering and yield characteristics i.e., number of flowers plant<sup>-1</sup>, number of setted pods plant<sup>-1</sup>, pods yield plant<sup>-1</sup>, weight of seeds pod<sup>-1</sup>, 100 seeds weight, weight of seeds (g) plant<sup>-1</sup> in the two growing seasons which were the most effective treatments in this respect compared with control and the other used treatments.

Generally, it could be recommended to spray common bean plants using growth stimulators such as yeast extract at 100 ml L<sup>-1</sup> or milagrow at 100 mg L<sup>-1</sup> to improve their growth, productivity and quality.

**Keywords:** Common bean, benzyladenine, milagrow, yeast and lithovit.

### INTRODUCTION

Common bean plant (*Phaseolus vulgaris* L.) is a member of legumes (Qin *et al.*, 2019). It is considered one of the most important vegetables, belonging to the family Fabaceae. Snap bean is an important economic vegetable crop grown in Egypt for local consumption and exportation. It is utilized for various purposes like fresh bean, dry pulses and edible podded type. It is rich in a variety of macro-and micro-nutrients, including starch (Punia *et al.*, 2020), the average crude protein content of common bean typically ranges from 21 to 25%, carbohydrate content varies between 60 and 65%, fibers content (soluble and insoluble) ranges between 3 and 7% , a very low amount of fat between 0.8 and 1.5% (Rathna and Manickavasagan, 2020.), minerals (P, Ca, K, Fe, and Mg) and vitamins (A, B1, B6, and B12) (Osorno *et al.*, 2020 and Mohamed *et al.*, 2023) and also source of antioxidant compounds such as phenols and ketones (Yang *et al.*, 2019), which have positive effects in managing diabetes, inhibiting obesity and increasing human metabolism (Liu *et al.*, 2020; Neil *et al.*, 2019; Nolan *et al.*, 2020). Egypt production of bean was 144809 tons from total area 90734 acre (Faostat, 2020).

Many investigators reported that improving physiological, morphological and anatomical performances of plants could be achieved by the application of different natural and chemical growth substances to enhance and maximize their growth, yield and seed quality characteristics as well.

In this respect, cytokinins are plant growth regulators that regulate plant growth, including cell division and leaf senescence (Gao, 2020). Cytokinins have been shown to have effects on many other physiological and development processes, including leaf senescence, formation and activity of shoot apical meristems, floral development, breaking of bud dormancy and seed germination. Cytokinins also appear to mediate many aspects of light-regulated development, including chloroplast differentiation, development of autotrophic metabolism, leaf and cotyledon expansion (Ikiba, 2017).

Benzyladenine (BA) is one of the cytokinins that improve quantitatively and/or qualitatively yield of many plants (Reda *et al.*, 2007). Application of benzyladenine is important for increasing plant height, number of leaves and fresh weight of leaves (Nahed and Aziz, 2007).

Milagrow is a natural growth promoter extracted from the pollen of cabbage flowers, it has great effectiveness in many field crops. Milagrow has combined effects of auxins, cytokines, gibberellins, ethylene and hydrogen cyanamid. The chemical composition of milagrow is 20% phosphorus, 10% potassium, 3% boron and 0.2% brassinolide (El-Emary and Abd El-Aal, 2018). Brassinolide is a natural plant growth promoter for all crops, which promotes growth, increases yield, improves quality and increases percentage of the fruit setting, spraying before flowering can promote the of flower buds and spraying during the flowering stage, can resist fruits dropping and flowers

dropping (Seadh *et al.*, 2012; Attia *et al.*, 2011). Milagrow (Brassinosteroids, BRs) has been found for stimulating cell division and elongation, differentiation of flowering buds, carbohydrate assimilation and activity of ATP, subsequently improving vegetative growth as well as enhancing physiological status then directing plant for earlier harvest and increasing fruit yield and quality features (Gabr *et al.*, 2011; Gomes *et al.*, 2006; Symons *et al.*, 2006). Brassinolide had a wide range of physiological and molecular responses in plants, such as stem elongation, pollen tube growth, leaf bending, photosynthesis, ethylene biosynthesis, proton pump activation, vascular differentiation, gene expression, nucleic acid and protein synthesis (Sasse, 2003).

As for yeast extract, it is a natural bio substance contains many nutrient elements and productive compounds of growth regulators like auxins, gibberellins and cytokinins. Dry yeast extract is a source of nutrients for plants because it contains amino acids, plant hormones, sugars, carbon, nitrogen, phosphorus, potassium, calcium, magnesium and micronutrients (El Sheikh *et al.*, 2022; Manea *et al.*, 2019). Yeast extract is a natural source of many growth substances (thiamine, riboflavin, niacin, pyridoxine and vitamins B1, B2, B3 and B12), cytokinins and many of the nutrient elements as well as organic compounds i.e., protein, carbohydrates, nucleic acid and lipids. Yeast is the best source of B-complex vitamin and a valuable source of bio-constituents, especially cytokinins (Amer, 2004 and El Sheikh *et al.*, 2022). It has been reported to be a rich source of phytohormones (especially cytokinins), vitamins, enzymes, amino acids and minerals (Khedr and Farid, 2000; Mahmoud, 2001). It was reported its stimulatory effects on cell division and enlargement, protein and nucleic acid synthesis and chlorophyll formation (Khater *et al.*, 2023). Improving growth and productivity of vegetable crops by the application of active yeast extract were recorded by (Khater *et al.*, 2023; Amer, 2004; El-Tohamy and El-Greadly 2007). It was additionally announced its stimulatory impact on cell division and expansion, protein and nucleic acid synthesis as well as chlorophyll formation (Khater *et al.*, 2023).

For lithovit compound, it is containing silica (5%), magnesium carbonate (4%) and calcium carbonate (75%), particles are extremely small, which gives them the ability to enter through the stomata of leaves with foliar spraying (Abo-Sedera *et al.*, 2016; Beinsan *et al.*, 2014). Lithovit (manufactured by Zeovita-GmbH, Berlin, Germany) and quantitatively analyzed by Wichmann and Basler (2006), who described it as a natural calcium carbonate foliar fertilizer supplemented with 12 different minerals that deliver fine particles (<10 µm) that can be easily adsorbed directly through the stomata of plant leaves. The positive effects of

lithovit on plant growth and chemical constituents were reported by Abu-Shleell (2017). Lithovit contains several elements or compounds, such as magnesium, which is the central element of the chlorophyll molecule; silica, which plays a vital role in increasing plant tolerance against abiotic stress, mainly concerning water relations and photosynthesis (Raven, 2003). The beneficial effect of this compound is being contains calcium carbonate (CaCO<sub>3</sub>) decomposes to calcium oxide (CaO) and carbon dioxide (CO<sub>2</sub>) in leaves stomata, and this CO<sub>2</sub> increases photosynthesis intensity, leading to increased carbon uptake and assimilation, there by increasing plant growth (Carmen *et al.*, 2014). Bilal (2010) lithovit is recommended by the European Community for organic farming according to EWG 2092/91. The micronutrients supplied with lithovit such as Mn, Cu, Zn, among others, influence plant physiology and metabolism and cell wall formation (Morsy *et al.*, 2018). Lithovit is a foliar fertilizer that improves plant growth and the yield of snap bean plant (Byan, 2014). Lithovit with the micronutrients provided an influence on cell wall formation, plant metabolism, and plant photosynthetic activity (Thorn and Rogan, 2015), that it works as an enzyme activator, a constituent of many enzymes, and a carrier of phosphorus in plant (Allison *et al.*, 2001; Haq and Mallarino, 2005).

So, this study aimed to investigate the effect of benzyladenine (BA), milagrow, yeast extract and lithovit foliar as spray treatments on growth, chemical compositions, anatomical features as well as yield characteristics of common bean plant (*Phaseolus vulgaris* L.).

## 2. MATERIAL AND METHODS

Two field experiments were conducted at the experimental Farm Station of Hort. Faculty of Agriculture Moshtohor, Benha University, Qalubia Governorate, Egypt, during seasons of 2022 and 2023. In order to study the effect of benzyladenine (BA), milagrow, yeast extract and lithovit on morphological, physiological, anatomical, flowering and yield characteristics of common bean (*Phaseolus vulgaris* L.).

Seeds of common bean (cv. Nebraska) were sown on the 1<sup>st</sup> of March during the 2022 and 2023 seasons. Three seeds were sown in hills on one side of the ridge at 15 cm space between the hills, 3 cm deep, after inoculation with root nodules bacteria (*Rhizobium phaseoli*). The standard inoculums contained about 270 spores g<sup>-1</sup> soil. The inoculums were placed below common bean seeds at sowing time in the soil. After complete germination plants were thinned into two plants per hill. The experimental design was a randomized complete block design (RCBD) with three replicates. Each plot consisted of three rows for each replicate (3m length X 0,7m width). This experiment

included 9 treatments. All field practices of growing common bean plants including fertilization, irrigation, manual weed control and so on were carried out as usual according to crop requirement recommendations.

#### The experimental treatments were as follows: -

Benzyladenine (BA) at 25 and 50 mg L<sup>-1</sup>, Milagrow at 50 and 100 mg L<sup>-1</sup>, Yeast extract at 50 and 100 ml L<sup>-1</sup> and Lithovit at 250 and 500 mg L<sup>-1</sup> as well as control (tap water).

Through plant growth and development, the growth substance treatments were applied as foliar spray three times starting at 25 days from sowing and repeated two times at 15-day intervals. The plants were sprayed with a hand pump mister. A surfactant (Tween 20) at a concentration of 0.01% was added to all tested solutions including the control. The foliar spraying solution was added to the point of runoff.

#### Treatments source:

1-Benzyladenine (BA) was exported from Algomhoria Chemical Company, Egypt.

2- Milagrow (Brassinosteroids, BRs) is a registered trademark product of Green India Co., India. It has been

obtained from Gaara Company, Cairo, Egypt. The compositions of milagrow are 0.2% brassinolide, 20% phosphorous, 10% potassium, and 3% boron (**Abou Elhassan et al., 2023; Seadh et al., 2012**)

3-Yeast extracts were prepared according to (**Abd El-Rahim et al., 1988**).

4- Lithovit as Nano particles (Ca, Mg and Fe nano or micro particles) containing calcium carbonate (80%), magnesium carbonate (4.6%) and Fe (0.75%) (**Carmen et al., 2014**) was obtained from Agro Link Company (3 Ibn- Eyas. St., Roxy, Heliopolis, Cairo).

The analysis of prepared yeast extract stock solution was: total protein (5.3%), total carbohydrates (4.7%), N (1.2%), P (0.13%), K (0.3%), Mg (0.013%), Ca (0.02%), Na (0.01%); micro-elements (mg L<sup>-1</sup>), Fe (0.13), Mn (0.07), Zn (0.04), Cu (0.04), B (0.016), Mo (0.0003), IAA (0.5 µg ml<sup>-1</sup>) and GA (0.3 µg ml<sup>-1</sup>).

Such analysis was according to (**Cotton, 1954**) for N, P, and K determination, the atomic absorption method of (**Nelson, 1944**) for mineral analysis, (**A.O.A.C. 2005**) for carbohydrate determination, the GLC method of (**Vogel, 1975**) for IAA and GAs determination and the method of (**Fletcher and McCullagh, 1986**) for cytokinins bioassay.

**Table (1): Physical and chemical properties of the experimental soil during 2022 and 2023 seasons were as follows.**

Soil analysis	Seasons	
	2022	2023
	Pre planting	Pre planting
<b>Soluble cations and anions (mmole L<sup>-1</sup>)</b>		
Ca <sup>++</sup>	1.3	2.6
Mg <sup>++</sup>	0.7	1.2
K <sup>+</sup>	0.4	0.7
Na <sup>+</sup>	5.1	8.3
Cl <sup>-</sup>	4.9	10.0
CO <sub>3</sub> <sup>2-</sup>	0.0	0.0
HCO <sub>3</sub> <sup>-</sup>	1.2	2.4
So <sub>4</sub> <sup>2-</sup>	1.4	0.4
<b>Chemical analysis</b>		
E.C* (dS m <sup>-1</sup> )	0.73	1.31
PH	7.92	7.78
<b>Particle size distribution (Mechanical analysis)</b>		
Course sand %	10.5	16.5
Find Sand %	16.1	10.0
Silt %	37.30	36.6
Clay %	36.1	36.9
Soil texture	Clay loam	Clay loam

**Sampling and collecting data: -****2.1. Morphological measurements: -**

Three plants from each replicate for each treatment were randomly taken at 55 days after sowing during both seasons and then separated into their organs and the following characteristics were recorded:

Plant height (cm), stem diameter (cm), number of branches plant<sup>-1</sup>, number of leaves plant<sup>-1</sup>, total leaves area plant<sup>-1</sup> by using disk methods according to (Derieux *et al.*, 1973), shoot fresh as well as dry weight (g) plant<sup>-1</sup>. The samples of the above ground i.e., vegetative parts were dried in the oven-dried for 48 h in 70°C to a constant weight and then the dry weight per plant was calculated. The dry samples were kept for chemical analysis.

**2.2. Photosynthetic pigments determination: -**

The photosynthetic pigments (chlorophyll a., b. and carotenoids) were extracted from fresh leaf samples (in the 4<sup>th</sup> apical leaves) at 55 days after sowing and determined using the methods described by (Fadeel, 1962; Wettstein, 1957).

**2.3. Anatomical features: -**

Specimens of the leaves were taken from the certain leaflet of the third apical leaf on the main stem of common bean plant at 55 days after sowing.

These vegetative specimens were killed and fixed in F.A.A. (10 ml formalin, 5 ml glacial acetic acid and 85 ml ethyl alcohol 70%), washed in 50% ethyl alcohol, dehydrated in a series of ethyl alcohols 70, 90, 95 and 100%, infiltrated in xylene, embedded in paraffin wax with a melting point 60-63°C, sectioned 15 microns in thickness for stem and 20 microns for the leaf (Sass, 1951), stained with the double stain method (Erythrosin and crystal violet), cleared in xylene and mounted in Canada balsam (Johanson, 1940). Stem and leaf sections were microscopically inspected to detect histological manifestations of noticeable responses resulting from treatments. Counts and measurements ( $\mu$ ) were taken using a micrometer eye piece. Averages of readings from 2 slides treatment<sup>-1</sup> were calculated.

**2.4. Endogenous phytohormones determination: -**

Endogenous phytohormones were quantitatively determined in common bean shoots at 55 days after sowing during the 2023 season. According to the method of Koshioka *et al.*, (1983) used for HPLC (High- Performance Liquid Chromatography) for the determination of auxin (IAA), gibberellic acid (GA<sub>3</sub>) and abscisic acid (ABA). Meanwhile, cytokinins were determined by HPLC according to Nicander *et al.*, (1993).

**2.5. Flowering stage measurements: -**

At the flowering stage, flowers number, flower setting percentage, flower abscission percentage, pods number plant<sup>-1</sup> were recorded and calculated.

**2.6. Dry pods yield and its components: -**

At harvest time (105 days after sowing in 2022 and 2023 seasons) three plants from each replicate were randomly taken to estimate the following characteristics: number of pods plant<sup>-1</sup>, number of seeds pod<sup>-1</sup>, pods weight (g) plant<sup>-1</sup>, seeds number plant<sup>-1</sup>, seeds yield (g) plant<sup>-1</sup>, weight of 100 seeds (g), pods yield (kg) fed<sup>-1</sup> and seeds yield (kg) fed<sup>-1</sup>.

**2.7. Chemical compositions: -**

Chemical analysis was carried out on the samples of dry shoots and seeds during the second season (2023).

**2.7.1. Determination of certain nutrient elements:**

The wet digestion of 0.2 g plant material with sulphuric and perchloric acids was carried out on shoots and dry seeds according to Piper (1947).

**Total nitrogen and crude protein:**

Total nitrogen was determined in the dry matter using microkjeldahl method as described by Horneck and Miller (1998), then calculated as a percentage of dry weight. Then, the crude protein was calculated according to the equation of A.O.A.C (2005).

Crude protein = Total nitrogen x 6.25

**Phosphorus:**

It was determined calorimetrically according to the method of Sandell (1950) and calculated as mg g<sup>-1</sup> dry weight.

**Potassium, calcium and sodium:**

They were or K, Ca and Na were determined by the flame photometer model Carl-Zeiss according to the method described by Horneck and Hanson (1998) and calculated as a percentage of dry weight.

**Iron and Manganese.**

Fe and Mn were determined using the atomic absorption spectrophotometer (Perkin Elmer 3110).

**2.7.2. Total carbohydrates :**

Total carbohydrates were determined in the dry shoot at 55 days after sowing and in the seeds at harvest time using the phenol-sulphuric acid method according to Dubois *et al.* (1954) and calculated as mg g<sup>-1</sup> dry weight.

**2.7.3. Determination of total sugars content: -**

Total sugars was determined according to the method described by (Kawamura *et al.*, 1966; Tanaka *et al.*, 1975 and Dubois *et al.*, 1956)

**2.8. Statistical analysis:**

Data of morphological, flowering and yield characteristics were statistically analyzed and the means were compared using the least significant differences (LSD) test at a 5% level of significance, according to Snedecore and Cochran (1989).

**3. RESULTS AND DISCUSSION****3.1. Effect of applied treatments on vegetative growth characteristics:**

Data in **Table 2** show that each of plant height (cm), number of branches plant<sup>-1</sup>, stem diameter (cm), number of leaves plant<sup>-1</sup> and total leaf area plant<sup>-1</sup> as well as shoot fresh and dry weights (g) plant<sup>-1</sup> of common bean plant were increased to reach the 5% level of significance with different applied treatments i.e., benzyladenine at 25 and 50 mg L<sup>-1</sup>, milagrow at 50 and 100 mg L<sup>-1</sup>, yeast extract at 50 and 100 ml L<sup>-1</sup> as well as lithovit at 250 and 500 mg L<sup>-1</sup> during the 2022 and 2023 seasons compared with the control.

The highest increases existed with yeast extract at 100 ml L<sup>-1</sup> followed by benzyladenine (BA) at 50 mg L<sup>-1</sup>, milagrow at 100 mg L<sup>-1</sup> and lithovit at 500 mg L<sup>-1</sup> compared with the control.

The profound effects of foliar-applied benzyladenine on enhancing plant growth parameters viz., plant height, number of branches and dry matter accumulation were reported by (**Samuel et al., 2000**). The promoting effects of cytokinins like benzyladenine (BA) on plant growth may be due to their stimulatory effects on cell division, cell elongation, nutrient mobilization, shoot initiation and growth, delay the processes of senescence and apical dominance alternation in plants (**Duck et al., 2004** and **Taiz and Zeiger, 2006**).

The promotive effect of milagrow on vegetative growth parameters may be due to the improvement of cell growth, differentiation, division and enlargement, alteration of membrane potential, and metabolism of nucleic acids and proteins (**Müssig, 2005; Dehghan et al., 2020; Abou Elhassan et al., 2023**). Milagrow (Brassinosteroids, BRs) has been found to stimulate cell division and elongation, flower bud differentiation, carbohydrate assimilation and ATP activity, subsequently improved vegetative growth, enhanced physiological status and directed trees to earlier harvest as well as increased fruit yield and quality (**Wang et al., 2004, Müssig, 2005, Gomes et al., 2006; Symons et al., 2006; Gabr et al., 2011**).

The enhancement effect of yeast extract may be due to its influence on metabolism, biological activity, photosynthetic pigments and enzyme activity which in turn encourage vegetative growth (**Wanas, 2002; El-Sherbeny et al., 2007** and **Adam et al., 2022**).

The micronutrients supplied with lithovit such as Mn, Cu, Z and among others, influence plant physiology and metabolism and cell wall formation (**Morsy et al., 2018**). Lithovit contains several elements or compounds, such as magnesium, which is the central element of the chlorophyll molecule and silica, which plays a vital role in increasing plant tolerance against abiotic stress, mainly concerning water relations and photosynthesis (**Raven, 2003**), calcium carbonate (CaCO<sub>3</sub>), which is decomposed into calcium oxide (CaO) and carbon dioxide (CO<sub>2</sub>) in the leaf stomata, where CO<sub>2</sub> increases

the intensity of photosynthesis (**Beinsan et al., 2014**). Moreover, **Ibrahim et al. (2016)** suggested that lithovit at this rate of feeding okra plant leaves with CO<sub>2</sub> gas from inside the leaves at a much higher rate than in the air, thus enhancing the basic process of photosynthesis and prompting assimilates accumulation, which is essential for building up protoplasm and protein, as well as inducing cell division, which resulted in an increase in cell number and cell size with an overall increase in the vegetative growth.

### **3.2. Effect of applied treatments on common bean bioconstituents:**

#### **3.2.1 Photosynthetic pigments:**

Data presented in Table 3 show that different applied treatments significantly increased the concentration of chlorophyll a, b and carotenoids in common bean leaves at 55 days after sowing during the 2022 and 2023 seasons compared with the control treatment. The maximum significant increase of these pigments existed with yeast extract at 100 ml L<sup>-1</sup> followed by benzyladenine at 50 mg L<sup>-1</sup> during the 2022 and 2023 seasons compared with the control treatment.

Herein, the yeast extract's activity as a bio-regulator may be responsible for the enhancement of photosynthetic pigments, because of its efficacy in the balance between photosynthesis and photorespiration in plants (**Olaiya, 2010**). Furthermore, yeast works to improve chlorophyll concentrations due to its rich contents of many essential elements, vitamins, and amino acids (**Abdelaal et al., 2017**). At the same time, yeast extract works on CO<sub>2</sub> release, which is reflected in the elevation of photosynthesis efficiency (**Khalil and Ismael, 2010** and **Mady, 2009**).

Benzyladenine is one of the most active synthetic cytokinins, influencing a wide range of plant developmental processes (i.e., chlorophyll synthesis and delayed leaf senescence) (**Müller and Sheen, 2007**). BA had a favorable effect on chlorophyll synthesis and accumulation in the leaves (**El-Maadawy et al., 2006**).

Brassinosteroids might activate or induce enzymes involved in chlorophyll biosynthesis or might surmount the stomatal limitations, thus escalating the CO<sub>2</sub> entry into the leaf and its availability for photosynthetic enzymes, resulting in elevated photosynthetic carbon fixing efficiency (**Holá et al., 2010**).

**Raven (2003)** and **Beinsan et al., (2014)** stated that lithovit contains several elements, such as magnesium, which is the central element of the chlorophyll molecule, and compounds such as calcium carbonate (CaCO<sub>3</sub>), which is decomposed to calcium oxide (CaO) and carbon dioxide (CO<sub>2</sub>) in the leaf stomata; hence, it leads to an increase in CO<sub>2</sub> levels within the plant leaf structure and by implication enhances photosynthetic efficiency.



Table (2) Effect of different applied treatments on some morphological characteristics of common bean (*Phaseolus vulgaris* L) plant at 55 days after sowing during the 2022 and 2023 seasons.

Treatment \ Trait	Plant height (cm)		Stem diameter (cm)		No. of leaves plant <sup>-1</sup>		Total leaf area (cm <sup>2</sup> ) plant <sup>-1</sup>		No. of branches plant <sup>-1</sup>		Shoot fresh weight (g) plant <sup>-1</sup>		Shoot dry weight (g) plant <sup>-1</sup>	
	2022	2023	2022	2023	2022	2023	2022	2023	2022	2023	2022	2023	2022	2023
benzyladenine at 25 mg L <sup>-1</sup>	44.67	47.33	0.77	0.83	31.00	33.33	3177.77	3498.50	7.33	7.67	171.27	169.83	25.18	24.97
benzyladenine at 50 mg L <sup>-1</sup>	47.33	51.00	0.60	0.67	37.00	38.00	4077.00	4180.27	8.67	8.33	188.80	189.57	27.19	27.31
milagrow at 50 mg L <sup>-1</sup>	48.00	38.00	0.47	0.47	25.67	26.67	2316.00	2400.77	5.00	5.33	166.23	159.40	24.77	23.76
milagrow at 100 mg L <sup>-1</sup>	50.33	40.33	0.50	0.40	28.33	30.33	2755.00	3064.00	6.33	6.00	174.70	174.20	25.51	25.43
yeast extract at 50 ml L <sup>-1</sup>	51.00	43.33	0.57	0.63	34.67	35.67	3707.27	3825.50	6.67	7.00	179.30	181.87	26.06	26.32
yeast extract at 100 ml L <sup>-1</sup>	53.67	45.00	0.83	0.87	40.33	40.33	4538.00	4546.77	8.00	7.67	198.83	197.43	28.44	28.24
lithovit at 250 mg L <sup>-1</sup>	41.67	35.33	0.40	0.53	23.00	25.00	2001.50	2158.50	4.33	4.67	162.63	155.77	24.39	23.35
lithovit at 500 mg L <sup>-1</sup>	43.33	37.00	0.47	0.60	27.33	28.00	2590.03	2717.03	6.00	5.67	168.50	163.53	24.95	24.29
control	37.00	33.00	0.37	0.53	20.00	23.00	1527.00	1720.50	3.67	4.00	156.53	151.17	23.63	22.82
L.S.D. at 5 %	1.82	2.47	0.09	0.09	2.28	2.91	252.90	262.1	0.99	1.22	4.06	3.05	0.57	0.42

Table (3) Effect of different applied foliar treatments on photosynthetic pigments (mg g<sup>-1</sup> F.W.) of common bean (*Phaseolus vulgaris* L) plant at 55 days after sowing during the 2022 and 2023 seasons.

Treatment \ Trait	Chlorophyll (a)		Chlorophyll (b)		Chlorophyll (a + b)		Carotenoids	
	2022	2023	2022	2023	2022	2023	2022	2023
benzyladenine at 25 mg L <sup>-1</sup>	0.706	0.540	0.350	0.109	1.056	0.650	0.289	0.279
benzyladenine at 50 mg L <sup>-1</sup>	0.894	1.026	0.376	0.313	1.270	1.339	0.327	0.439
milagrow at 50 mg L <sup>-1</sup>	0.603	0.741	0.247	0.150	0.850	0.892	0.219	0.324
milagrow at 100 mg L <sup>-1</sup>	0.872	0.827	0.276	0.167	1.149	0.994	0.376	0.398
yeast extract at 50 ml L <sup>-1</sup>	0.376	0.930	0.140	0.270	0.516	1.200	0.173	0.384
yeast extract at 100 ml L <sup>-1</sup>	1.078	1.133	0.482	0.378	1.560	1.511	0.382	0.478
lithovit at 250 mg L <sup>-1</sup>	0.695	0.641	0.224	0.098	0.920	0.739	0.272	0.368
lithovit at 500 mg L <sup>-1</sup>	0.709	0.712	0.270	0.158	0.98	0.871	0.270	0.342
control	0.305	0.382	0.069	0.064	0.374	0.446	0.161	0.232

**Table (4)** Effect of different applied foliar treatments on some chemical constituents level of common bean shoot (*Phaseolus vulgaris* L.) at 55 days after sowing during the 2023 season.

Trait Treatment	N %	P %	K %	Ca %	Na %	Fe mgL <sup>-1</sup>	Mn mgL <sup>-1</sup>	Protein %	Total carbohydrates %	Total sugars %
benzyladenine at 25 mg L <sup>-1</sup>	1.91	0.51	2.99	2.43	1.23	0.33	1.00	11.98	54.91	7.19
benzyladenine at 50 mg L <sup>-1</sup>	2.38	0.65	2.53	3.10	1.68	0.32	1.20	14.88	54.32	7.68
milagrow at 50 mg L <sup>-1</sup>	2.80	0.50	2.60	3.47	1.20	0.35	1.62	17.50	55.17	7.80
milagrow at 100 mg L <sup>-1</sup>	2.80	0.69	2.72	3.27	1.11	0.49	0.60	17.50	56.48	7.73
yeast extract at 50 ml L <sup>-1</sup>	2.11	0.57	2.79	3.39	1.24	0.50	0.62	13.19	55.41	7.45
yeast extract at 100 ml L <sup>-1</sup>	2.18	0.53	2.72	2.78	1.49	0.44	0.60	13.66	55.90	7.32
lithovit at 250 mg L <sup>-1</sup>	2.42	0.51	2.56	2.53	1.35	0.61	0.62	15.16	54.47	7.39
lithovit at 500 mg L <sup>-1</sup>	2.62	0.48	2.60	2.69	1.46	1.02	1.20	16.38	54.02	7.67
control	1.98	0.43	2.38	1.94	1.08	0.31	0.53	12.38	53.76	7.18

### **3.2.2. minerals, crud protein, total carbohydrate and total sugars:**

Data in Table 4 illustrate that all applied foliar treatments effectively increased the of N, P, K, Ca, Na, Fe, Mn, crude protein, total carbohydrate and total sugars in the shoots of treated plants compared with those of the control during the 2023 season. In this respect the most effective treatments were milagrow at 100 mg L<sup>-1</sup> and 50 mg L<sup>-1</sup> followed by yeast extract with its two concentrations, benzyladenine at 50 mg L<sup>-1</sup> and lithovit at 500 mg L<sup>-1</sup>, respectively.

In this respect, **El-Emary and Abd El-Aal (2018)** Indicated that foliar spraying of milagrow significantly increased the percentage of nitrogen, crude protein, phosphorus and potassium in treated tomato plants compared with the untreated plants. The increase may be due to milagrow being a natural growth promoter. Milagrow is a rich source of auxins, cytokines, gibberellins, ethylene and hydrogen cyanamid and its chemical composition is 20% phosphorus, 10% potassium, 3% boron and 0.2% brassinolide. Milagrow stimulate chemical compositions biosynthesis activity by the combined effects of auxins, cytokines, gibberellins, ethylene and hydrogen cyanamid (**Seadh et al., (2012)**)

Yeast extract may increase absorption of different elements by roots and also their translocation and accumulation in leaves (**Hammad, 2008; Mady, 2009**). The application of yeast caused a significant increase of N, P and K% in pea leaves which could be attributed to its minerals, carbohydrates and hormonal contents (**Hammad, 2008**). **Taha et al., (2016)** mentioned that yeast also facilitates the growth of plants by improving the uptake of nutrients and production of some phytohormones.

These results of benzyladenine conform with those obtained by **Zewail et al., (2019)** on soybean plants. Also, **Mahmoud et al., (2016)** reported that foliar application with benzyladenine significantly increased lupine plant chemical composition.

Lithovit contains several elements, such as magnesium, which is the central element of the chlorophyll molecule, and compounds such as calcium carbonate (CaCO<sub>3</sub>), which is decomposed to calcium oxide (CaO) and carbon dioxide (CO<sub>2</sub>) in the leaf stomata; hence, it leads to increased CO<sub>2</sub> levels within the plant leaf structure and, by implication, enhances photosynthetic efficiency (**Beinsan et al., 2014**). Consequently, chemical bioconstituents increased.

### **3.3. Endogenous phytohormones:**

Data in **Table (5)** show changes in the levels of endogenous phytohormones i.e., gibberellins, auxins, cytokinins and abscisic acid, in common bean shoots treated with benzyladenine at 50 mg L<sup>-1</sup>, milagrow at 100 mg L<sup>-1</sup>, yeast extract at 100 ml L<sup>-1</sup> and lithovit at 500 mg L<sup>-1</sup> (the most effective treatments, which greatly improved the morphological, metabolic performances of common bean plant as previously mentioned) and control at 55 days after sowing during the 2023 season.

Concerning gibberellin, treatment of benzyladenine at 50 mg L<sup>-1</sup> achieved the highest value followed by milagrow at 100 mg L<sup>-1</sup> then lithovit at 500 mg L<sup>-1</sup>, meanwhile yeast extract at 100 ml L<sup>-1</sup> gave the lowest value compared with the control.

As for auxin level, benzyladenine at 50 mg L<sup>-1</sup> was the most effective treatment followed by lithovit at 500 mg L<sup>-1</sup> then yeast extract at 100 ml L<sup>-1</sup> and the control, respectively, while milagrow at 100 mg L<sup>-1</sup> ranked last in this respect.

For cytokinins, Lithovit at 500 mg L<sup>-1</sup> gave the highest value compared with the control followed by



benzyladenine at 50 mg L<sup>-1</sup>, milagrow at 100 mg L<sup>-1</sup> and yeast extract at 100 ml L<sup>-1</sup>, respectively.

Generally, phytohormones that promote growth aspects, i.e., growth promoters (gibberellins, auxins and cytokinins), levels in common bean shoots were increased with the assigned treatments during the 2023 season compared with the control. Here, lithovit at 500 mg L<sup>-1</sup> gave the highest value of total promoters followed by benzyladenine at 50 mg L<sup>-1</sup>, milagrow at 100 mg L<sup>-1</sup> and yeast extract at 100 ml L<sup>-1</sup> in descending order. Concerning the growth inhibitor, (abscisic acid) its level was reduced with various assigned treatments during 2023 season compared with the control, but the reduction was more obvious with lithovit at 500 mg L<sup>-1</sup>. These increases in the endogenous growth promoters and decreases in growth inhibitor substances in response to applied treatments may be attributed to their effect on enhancing the biosynthesis of growth promoters and decreasing the biosynthesis and action of growth inhibitors (abscisic acid).

**Abd El-Aal and Eid-Rania (2018)** indicated that application of lithovit at a rate of 500 mg L<sup>-1</sup> on soybean gave the highest values of phytohormones concentration.

Cytokinins are phytohormones influencing a wide range of plant developmental processes for example shoot branching and flowering (**Buban, 2000** and **Müller and Sheen, 2007**). Foliar application with benzyladenine significantly increased endogenous phytohormones concentration in the shoots of soybean plant (**Zewail et al., 2019**).

Milagrow is a natural growth promoter, it has combined effects of auxins, cytokines, gibberellins, ethylene and hydrogen cyanamid (**El-Emary and Abd El-Aal, 2018; Seadh et al., 2012; Attia et al., 2011**). Spraying tomato plants with milagrow caused an increase in phytohormones i.e., gibberellins, auxins, cytokinins and salicylic acid whereas reduced abscisic acid level (**El-Emary and Abd El-Aal, 2018**).

Application of yeast extract acts as a biostimulents that improves growth and yield of different crops and it has beneficial roles due to its content of biologically active substances such as phytohormones, different nutrients, protein, amino acids and vitamin B (**Dawood**

*et al., 2013; Abbas 2013; Mahmoud et al., 2016; Abdelaal et al., 2017; Abdallah 2020*). **El-Shafey et al., (2016)** Yeast extract foliar spray at 5 g L<sup>-1</sup> and 10 g L<sup>-1</sup> increased endogenous phytohormones such as (cytokinin, GA<sub>3</sub> and IAA) of faba bean plant.

### **3.4. Effect of applied treatments on the anatomical characteristics of common bean:**

#### **3.4.2. Effect of applied treatments on leaf anatomy:**

Data in Table 6 and Fig.1 indicate that most studied leaf anatomical features were increased with different applied treatments compared with the control. Among these anatomical features were the most important ones, i.e., the thickness of each midrib, lamina, palisade tissue, and spongy tissue, as well as the length and width of the vascular bundle, phloem, and xylem tissues, in addition to number of xylem vessels in vascular bundle.

These increases were more obvious with yeast extract at 50 ml L<sup>-1</sup> and 100 ml L<sup>-1</sup> followed by benzyladenine at 25 mg L<sup>-1</sup> and 50 mg L<sup>-1</sup> as well as milagrow at 50 mg L<sup>-1</sup>.

Foliar application with the most effective concentration of active yeast extract (ml L<sup>-1</sup>) on kidney bean 'Giza 6', increased the thickness of both midvein and lamina of the leaflet due mainly to increments in the thickness of palisade and spongy tissues as well as the main vascular bundle of the midvein (**Nassar et al., 2011**).

The positive effects of benzyladenine on different anatomical traits of stems and leaf blades might be due to its ameliorative effect on endogenous cytokinins (**Gad-Mervat, 2005**).

**Hayat et al., (2003)** revealed that brassinosteroids regulate metabolism through the signals of auxins for promoting cell division and xylem differentiation. The positive effects of milagrow may be due to the improvement of cell growth, differentiation, division and enlargement (**Müssig, 2005; Dehghan et al., 2020**). Lithovit caused an increase in the thickness of the midvein region, the number of vessels in the midvein bundle and the average diameter of the vessel (**Nawar et al., 2018**). Increasing of leaf anatomy characters may be positively correlated with increasing cell division and elongation (**El-Emary et al., 2018**).

Table (5) Effect of applied treatments on endogenous phytohormones concentration or level in common bean (*Phaseolus vulgaris* L) shoots at 55 days after sowing during the 2023 season.

Plant hormones Treatments	Gibberellins $\mu\text{g g}^{-1}$ F.w.	Auxins $\mu\text{g g}^{-1}$ F.w.	Cytokinins $\mu\text{g g}^{-1}$ F.w	Total Promoters $\mu\text{g g}^{-1}$ F.w	Abscisic acid $\mu\text{g g}^{-1}$ F.w
benzyl adenine at 50 mg L <sup>-1</sup>	569.8254	48.4463	449.9522	1068.2239	4.3580
milagrow at 100 mg L <sup>-1</sup>	544.3310	25.7963	424.8314	994.9587	1.9734
yeast extract at 100 ml L <sup>-1</sup>	450.0930	37.1409	392.3918	879.6257	3.7936
lithovit at 500 mg L <sup>-1</sup>	526.1633	44.9678	591.6128	1162.7439	1.5455
Control	519.2139	34.5010	262.7003	816.4152	21.5068

Table (6) Effect of applied treatments on the mean counts and measurements of certain anatomical features of common bean (*Phaseolus vulgaris* L) leaflet at 55 days after sowing during the 2023 season.

Treatments Characteristics	benzyladenine at 25 mg L <sup>-1</sup>	benzyladenine at 50 mg L <sup>-1</sup>	milagrow at 50 mg L <sup>-1</sup>	milagrow at 100 mg L <sup>-1</sup>	yeast extract at 50 ml L <sup>-1</sup>	yeast extract at 100 ml L <sup>-1</sup>	lithovit at 250 mg L <sup>-1</sup>	lithovit at 500 mg L <sup>-1</sup>	Control
Thickness of leaf midrib.	1449.0	1449.0	1746.0	1620.0	1395.0	1593.0	1260.0	1305.0	828.0
Thickness of lamina.	364.95	286.2	396.9	272.7	330.2	275.4	299.7	343.8	238.5
Upper epidermis thickness.	32.85	31.50	33.30	31.50	31.50	26.10	27.00	33.30	25.20
Lower epidermis thickness.	25.20	17.10	27.00	25.20	25.20	18.00	22.50	27.00	18.90
Palisade tissue thickness.	144.0	133.2	189.0	108.0	117.0	117.0	108.0	135.0	90.0
Spongy tissue thickness.	128.7	97.2	117.0	108.0	117.0	90.0	108.0	108.0	81.0
Length of the largest v. bundle	392.3	407.7	497.7	432.0	549.0	450.0	405.0	422.0	171.0
Thickness of phloem.	96.3	53.1	74.7	72.0	90.0	90.0	90.0	71.1	45.0
Thickness of xylem	288.0	261.9	306.0	270.0	378.0	288.0	225.0	270.0	126.0
No. of xylem rows in vascular bundle.	6.0	9.0	6.0	5.0	5.0	7.0	9.0	7.0	4.0
Diameter of the widest xylem vessel.	79.2	52.2	63.0	51.3	60.3	59.4	57.6	49.5	36.9

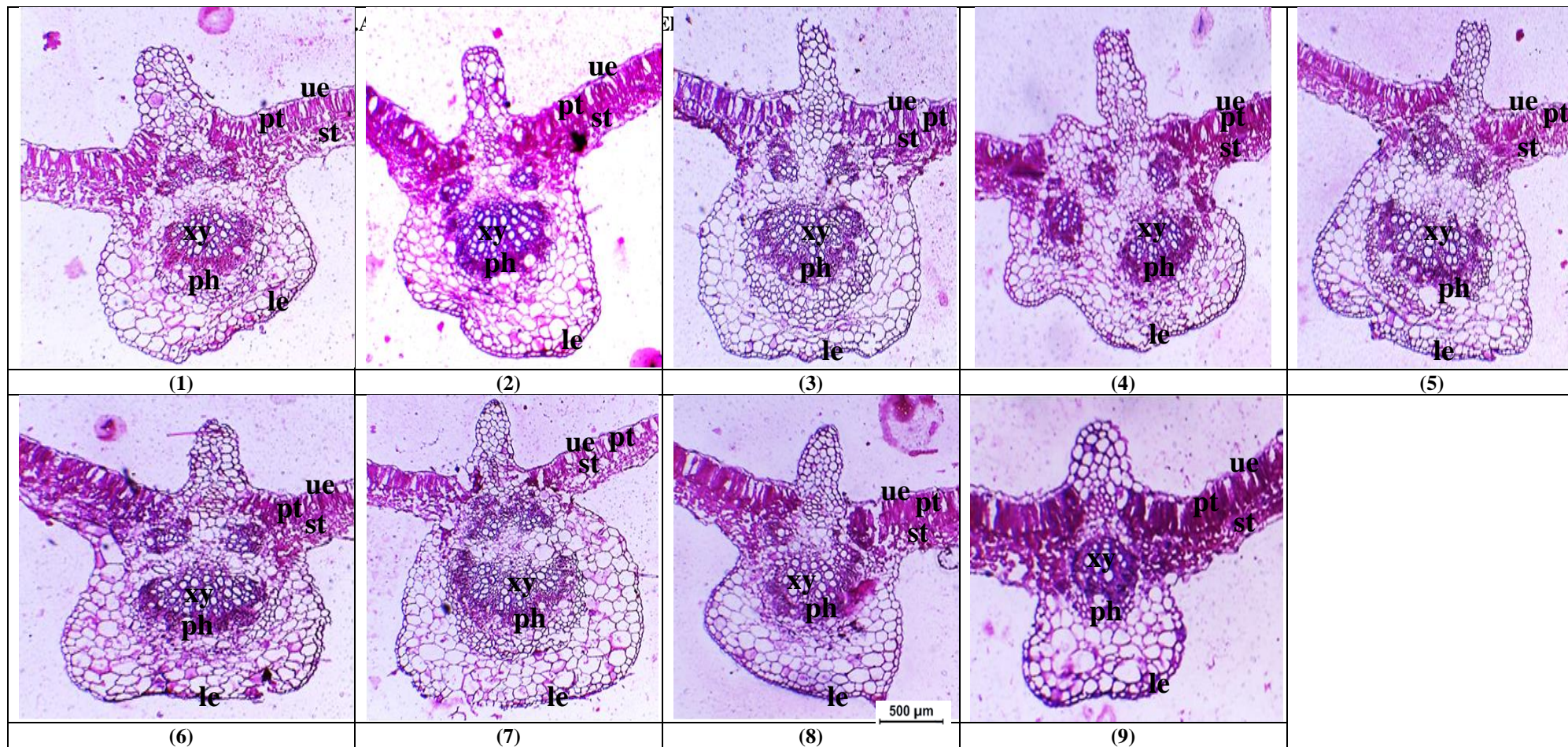


Fig. (1): Transverse sections (X = 40) through the terminal leaflet of the 4<sup>th</sup> apical leaf of common bean plant at 55 days after sowing as affected by different applied treatments.

Where: (1): benzyladenine at 25 mg L<sup>-1</sup> (2): benzyladenine at 50 mg L<sup>-1</sup> (3): milagrow at 50 mg L<sup>-1</sup> (4): milagrow at 100 mg L<sup>-1</sup>  
 (5): yeast extract at 50 ml L<sup>-1</sup> (6): yeast extract at 100 ml L<sup>-1</sup> (7): lithovit at 250 mg L<sup>-1</sup> (8): lithovit at 500 mg L<sup>-1</sup> (9): Control  
 ue = upper epidermis le = lower epidermis pt = palisade tissue st = spongy tissue ph = phloem tissue xy = xylem tissue

### **3.5. Effect of applied treatments on flowering characteristics:-**

Data in Table 7 as shown in Table 8, different applied spraying treatments had ameliorative effects on flowering characteristics of common bean plants since they caused a significant increase in the number of pods plant<sup>-1</sup> compared with the untreated plants during both seasons of this study. This increase was accompanied by a significant increase in the number of flowers and the percentage of flower set as well as a significant reduction in the flower abscission. In this respect the most efficacious treatment was yeast extract at 100 ml L<sup>-1</sup> followed by milagrow at 100 mg L<sup>-1</sup> then lithovit at 250 mg L<sup>-1</sup> or benzyladenine at 50 mg L<sup>-1</sup>.

Improving the productivity of some vegetable plants by using a spray application with yeast extract was indicated by **Abou EL-Yazied and Mady (2012); Kamal and Ghanem, (2012); Mahmoud et al., (2013); Zewail et al., (2019)**, who attributed that to its bio-substances contents, such as phytohormones, especially cytokinines, sugars, vitamins, enzymes, amino acids and minerals which reflected in flowering improvement.

Milagrow (Brassinosteroids, BRs) has been found to stimulate flower bud differentiation (**Wang et al., 2004; Mussig, 2005; Gomes et al., 2006; Symons et al., 2006; Gabr et al., 2011**). Milagrow is a natural growth promoter, increases fruit setting percentage, promotes flowering buds formation and resists flowers and fruits dropping (**El-Emary and Abd El-Aal, 2018**).

Application of lithovit caused an increase in photosynthetic efficiency which reflected in flowering improvement (**Abass et al., 2022; Abd El-baset, 2018**).

### **3.6. Effect of applied treatments on yield and its components:**

Data in Table 8 show the effect of benzyladenine at 25 mg L<sup>-1</sup> and 50 mg L<sup>-1</sup>, milagrow at 50 mg L<sup>-1</sup> and 100 mg L<sup>-1</sup>, yeast extract at 50 ml L<sup>-1</sup> and 100 ml L<sup>-1</sup>, and lithovit at 250 mg L<sup>-1</sup> and 500 mg L<sup>-1</sup> applied as foliar spray treatments on yield characteristics (i.e., No. of pods plant<sup>-1</sup>, pods weight (g) plant<sup>-1</sup>, No. of seeds pod<sup>-1</sup>, No. of seeds plant<sup>-1</sup>, seeds weight (g) plant<sup>-1</sup>, 100-seeds weight (g), pods yield fed<sup>-1</sup> (kg) and seed yield fed<sup>-1</sup> (kg) ) at harvest time during the 2022 and 2023 seasons.

Data indicate that different applied treatments increased yield traits during the 2022 and 2023 seasons

compared with the control treatment. Yeast extract at 100 ml L<sup>-1</sup> followed by milagrow at 100 mg L<sup>-1</sup> gave the highest significance increases, respectively when compared with the control and other treatments in both seasons.

The enhancing effect of such treatments on common bean plant (*Phaseolus vulgaris* L.) yield components may be due to their ameliorative effects on flowering and setting percentage, consequently the number of pods and their weight. Also, this could be due to the promotional effects of the same treatments on growth behavior, metabolic activity levels, mineral content and phytohormones level and through increasing the assimilates and their translocations from leaves to the developing seeds in pods all of them positively correlated with yield quantity and quality.

The present results are generally in agreement with those reported by (**Zewail, 2014; El-Shafey et al., 2016; Muhammad et al., 2020**) for yeast; (**Ayad and Gamal El-Din, 2011; Mahmoud et al., 2016; Zewail et al., 2019 ; Shyaa and Kisko, 2023**) for benzyladenine (BA); (**El-Bassiony et al., 2012; Netwal et al., 2018; Furio et al., 2022**) for milagrow (Brassinosteroids); (**Abd El-Aal and Eid-Rania, 2018; Byan, 2020 ; Saad et al., 2021**) for lithovit.

### **3.7. Effect of applied treatments on chemical constituents of common bean seeds:**

As shown in Table 9 different foliar spraying treatments caused an obvious increases in the concentration all determined chemical constituents of common bean seeds, mainly nitrogen, phosphorus, potassium, calcium, sodium, iron, manganese as well as crude protein and total carbohydrate in addition to total sugars % compared with the control during the 2023 season.

In this respect, the most effective treatments were yeast extract at 100 ml L<sup>-1</sup> followed by benzyladenine at 50 mg L<sup>-1</sup> then lithovit at 500 mg L<sup>-1</sup> and milagrow at 100 mg L<sup>-1</sup>, respectively compared with the control.

Other studies also, reported nearly similar results **El-Shafey et al., 2016 and Muhammad et al., 2020**) for yeast; **Shyaa and Kisko, 2023**) for benzyladenine ; **Netwal et al., 2018 and Furio et al., 2022**) for milagrow (Brassinosteroids); (**Abd El-Aal and Eid-Rania, 2018 and Byan, 2020**) for lithovit

**Table (7)** Effect of applied treatments on flowering characteristics of common bean (*Phaseolus vulgaris* L) plant at flowering stage during the 2022 and 2023 seasons.

Trait Treatments	No. of flowers plant <sup>-1</sup>		No. of pods plant <sup>-1</sup>		Flowers setting (%)		Flowers abscission %	
	2022	2023	2022	2023	2022	2023	2022	2023
benzyladenine at 25 mg L <sup>-1</sup>	68.67	73.00	31.67	36.00	46.09	49.31	53.91	50.69
benzyladenine at 50 mg L <sup>-1</sup>	64.33	65.67	41.33	43.00	64.26	65.53	35.74	34.47
milagrow at 50 mg L <sup>-1</sup>	66.33	70.00	45.00	48.00	67.84	68.57	32.16	31.43
milagrow at 100 mg L <sup>-1</sup>	73.33	77.00	51.00	53.67	69.53	69.72	30.47	30.28
yeast extract at 50 ml L <sup>-1</sup>	68.00	72.00	48.00	51.00	70.57	70.83	29.43	29.17
yeast extract at 100 ml L <sup>-1</sup>	74.67	79.00	53.33	56.33	71.39	71.34	28.61	28.66
lithovit at 250 mg L <sup>-1</sup>	70.33	75.33	34.33	40.33	48.76	53.55	51.24	46.45
lithovit at 500 mg L <sup>-1</sup>	65.67	68.33	43.33	46.33	65.98	67.83	34.02	32.17
Control	59.67	62.00	25.33	29.00	42.39	46.76	57.61	53.24
L.S.D. at 5 %	2.15	1.52	3.09	1.83	3.28	2.84	3.28	2.84

**Table (8)** Effect of different applied treatments on yield and yield components of common bean (*Phaseolus vulgaris* L) plant at harvest time during the 2022 and 2023 season.

Trait Treatment	Pods weight (g) plant <sup>-1</sup>		No. of seeds pods <sup>-1</sup>		No. of seeds plant <sup>-1</sup>		seeds weight (g) plant <sup>-1</sup>		100-seeds weight (g)		Pods yield fed <sup>-1</sup> (kg)		Seeds yield fed <sup>-1</sup> (kg)	
	2022	2023	2022	2023	2022	2023	2022	2023	2022	2023	2022	2023	2022	2023
benzyladenine at 25 mg L <sup>-1</sup>	87.63	94.47	3.91	3.78	123.33	136.00	36.70	45.74	29.75	33.63	1641.7	1808.3	978.6	1093.0
benzyladenine at 50 mg L <sup>-1</sup>	95.43	102.73	3.79	3.52	132.67	144.33	41.58	51.87	31.35	35.93	1758.3	1941.7	1116.4	1259.4
milagrow at 50 mg L <sup>-1</sup>	100.73	114.10	3.27	3.19	147.00	153.00	48.09	57.72	32.72	37.74	1950.0	2100.0	1281.8	1401.0
milagrow at 100 mg L <sup>-1</sup>	108.13	121.57	3.21	3.33	159.33	179.33	58.47	71.07	36.69	39.64	2141.7	2333.3	1480.2	1562.2
yeast extract at 50 ml L <sup>-1</sup>	102.77	118.67	3.13	3.34	154.00	170.00	52.94	65.91	34.40	38.79	2033.3	2275.0	1375.7	1562.2
yeast extract at 100 ml L <sup>-1</sup>	112.37	127.93	3.20	3.31	170.33	186.00	63.47	76.90	37.26	41.35	2225.0	2416.7	1584.8	1753.8
lithovit at 250 mg L <sup>-1</sup>	92.77	98.67	3.21	3.36	129.67	141.67	40.18	49.05	31.00	34.62	1700.0	1875.0	1056.7	1186.2
lithovit at 500 mg L <sup>-1</sup>	97.97	109.23	3.25	3.26	140.67	150.67	45.87	55.24	32.62	36.66	1891.7	2008.3	1222.0	1320.0
control	83.27	89.47	4.12	3.84	103.67	111.33	30.17	34.80	29.09	31.25	1358.3	1366.7	855.4	872.7
L.S.D. at 5 %	3.37	4.09	N.S.	N.S.	4.38	5.40	2.39	3.23	1.90	2.05	91.8	88.6	55.5	50.8

**Table (9)** Effect of applied treatments on some bioconstituents of common bean (*Phaseolus vulgaris* L) seeds at harvest time during 2023 season.

Trait Treatment	N %	P %	K %	Ca %	Na %	Fe mg g <sup>-1</sup> D.W	Mn mg g <sup>-1</sup> 1 D.W	Protein %	Carbohydrates %	Total sugars %
benzyladenine at 25 mg L <sup>-1</sup>	2.22	0.72	1.88	1.12	0.67	0.56	0.77	13.88	63.81	9.48
benzyladenine at 50 mg L <sup>-1</sup>	2.8	0.73	1.69	1.13	0.82	0.58	0.72	17.5	62.35	9.46
milagrow at 50 mg L <sup>-1</sup>	2.55	0.70	1.78	1.28	0.89	0.50	0.77	15.94	65.53	9.35
milagrow at 100 mg L <sup>-1</sup>	2.55	0.69	1.56	1.13	0.78	0.72	0.88	15.94	64.42	9.64
yeast extract at 50 ml L <sup>-1</sup>	2.82	0.72	1.67	1.41	0.93	0.62	0.82	17.66	63.71	9.86
yeast extract at 100 ml L <sup>-1</sup>	2.82	0.71	1.71	1.36	0.74	0.60	0.62	17.66	65.34	9.87
lithovit at 250 mg L <sup>-1</sup>	2.82	0.68	1.57	1.61	0.93	0.89	0.99	17.66	64.49	9.08
lithovit at 500 mg L <sup>-1</sup>	2.66	0.66	1.53	1.32	0.56	0.92	0.92	16.66	63.46	9.79
control	2.12	0.62	1.48	1.04	0.49	0.93	0.73	13.25	61.27	8.98

## REFERENCES

- [1] **A.O.A.C. (2005)**: Official methods of analysis, 18<sup>th</sup> ed., Association of Official Agricultural Chemists, Washington, DC, USA.
- [2] **Abass, M. M; Essa, R. S. T. and Fahmy, A. A. (2022)**: Impact Of Potassium Fertilizer And Lithovit On Growth, Flowering And Chemical Constituents Of Arabian Jasmine Var. Double Petals Plant. Sinai Journal of Applied Sciences, 11(6):1115-1128.
- [3] **Abbas, S. M. (2013)**: The influence of biostimulants on the growth and on the biochemical composition of *Vicia faba* cv. Giza 3 beans. Romanian Biotechnological Letters, 18(2): 8061-8068.
- [4] **Abd El-Aal, M. M. M. and Eid-Rania, S. M. (2018)**: Effect of foliar spray with lithovit and amino acids on growth, bioconstituents, anatomical and yield features of soybean plant. Annals of Agricultural Science, Moshtohor, 56(4<sup>th</sup> ICBA): 187-202.
- [5] **Abd El-baset, M. M. (2018)**: Effect of Nano Material Lithovit on Growth, Flowering and Chemical Composition of *Echinacea purpurea* (L.). Journal of Plant Production, 9(6): 531-536.
- [6] **Abdallah, H.R. (2020)**: Effect of foliar application with urea, benzyladenine and dry yeast on flowering and fruiting of Minneola Tangelo trees to reduce the severity of alternate bearing phenomena. Middle East J. Agric. Res., 09(04), 893-933
- [7] **Abdelaal, K.A; Hafez, Y.M; El Sabagh, A. and Saneoka, H. (2017)**: Ameliorative effects of abscisic acid and yeast on morpho-physiological and yield characteristics of maize plant (*Zea mays* L.) under water deficit conditions. Fresenius Environ. Bull., 26, 7372–7383
- [8] **Abo-Sedera, F.A; Shams, A.S; Mohamed, M.H.M. and Hamoda, A. H.M. (2016)**: Effect of organic fertilizer and foliar spray with some safety compounds on gridgeth and productivity of snap bean. Ann. Agric. Sci., Moshtohor, 54 (1): 105 – 118
- [9] **Abou Elhassan, M. H; Bosila, H. A; Hamza, M. A; Elateeq, A. A; Abdel-Gawad, A. I. M. and Soliman, M. N. A. (2023)**: Seaweed extract and brassinolide can enhance the dwarfing characteristics of potted chrysanthemum. International Journal of Agricultural Technology 19(3):849-876.
- [10] **Abou EL-Yazied, A. and Mady, M.A. (2012)**: Effect of boron and yeast extract foliar application on growth, pod setting and both green pod and seed yield of broad bean (*Vicia faba* L.). Journal of Applied Sciences Research, 8(2): 1240- 1251.
- [11] **Abu-Shleell, M. K. K. M. (2017)**: Botanical studies on Moringa plants. M.Sc. Thesis, Fac. Of Agric., Benha Univ., Egypt.
- [12] **Adam, A. R; Allafe, M. A and Omar, E. A. (2022)**: Biostimulants Influence (Licorice and Yeast Extract) on Vegetative Growth of Faba Bean (*Vicia faba* L.). Journal of Plant Production, 13(7), 321-324.
- [13] **Allison, M.F; Fower, J.H. and Allen, E.J. (2001)**: Factors affecting the magnesium nutrition of potatoes (*Solanum tuberosum* L.) J. Agric. Sci. Cambridge, 137:397- 409. Applied Microbiology and Biotechnology, 102(19): 8229-8259.
- [14] **Amer, S.S.A. (2004)**: Growth, green pods yield and seeds yield of common bean (*Phaseolus vulgaris* L.) as affected by active dry yeast, salicylic acid and their interaction. J. Agric. Sci., Mansoura Univ., 29(3): 1407- 1422.
- [15] **Attia, A. N. E; Badawi, M. A; Seadh, S. E. and El-Hety, S. M. S. (2011)**: "Response of growth and petal yield of safflower to sowing dates, nitrogen fertilizer, levels and times of foliar application with natural growth promoters "Melagrow"", Mansoura Journal of Plant Production, Vol. 2 No. 2, pp. 1703–1716.
- [16] **Ayad, H.S and Gamal El-Din, K.M. (2011)**: Effect of atonik and benzyladenine on growth and some biochemical constituents of lupine plant (*Lupinus termis* L) American Eurasian. J. Agric. & Environ. Sci., 10(4): 519-524.
- [17] **Beinsan, C; Şumalan, R; Gâdea, Ş. and Vâtcă, S. (2014)**: Physiological indicators study involved in productivity increasing in tomato. ProEnvironment Promediu, 7(20).
- [18] **Bilal, B.A. (2010)**: Lithovit®: An innovative fertilizer. The 3<sup>rd</sup> e Conference on Agricultural Biosciences (IeCAB 2010), 1st- 15<sup>th</sup> June 2010. <http://www.slideserve.com/madison/lithovitan->
- [19] **Buban, T. (2000)**: The use of benzyl adenine in orchard fruit growing: A mini review. Plant Growth Reg., 32,381-390.
- [20] **Byan, U. A. (2020)**: Effect of foliar spray with aqueous extract of date palm pollen grains and Lithovit on common bean plants under different irrigation levels. Zagazig Journal of Agricultural Research, 47(3), 677-691.
- [21] **Byan, U. A. I. (2014)**: Influence of using some safety materials on water requirement and water use efficiency of snap bean plant. Arab Univ. J. Agric. Sci., Ain Shams Univ., Cairo, 22 (2): 381-394.
- [22] **Carmen, B; Sumalan, R.; Gadea, S. and Vatca, S. (2014)**: Physiological indicators study involved in productivity increasing in tomato. Pro-environment, 7: 218 – 224.

- [23] **Cotton, R. H. (1954):** Determination of nitrogen, phosphorus and potassium in leaf tissue, application of micro methods. *Ind. and Eng. Chem. Anal. Edit.*, 17 (11):734-738.
- [24] **Dawood, M. G; El-Lethy, S. R; Sadak, M.S. (2013):** Role of methanol and yeast in improving growth, yield, nutritive value and antioxidants of soybean. *World Appl, Sci, J.*, 26: 06–14
- [25] **Dehghan, M., Balouchi, H., Yadavi, A. and Zare, E. (2020):** Improve wheat (*Triticum aestivum*) performance by brassinolide application under different irrigation regimes. *South African Journal of Botany*, 130:259-267.
- [26] **Derieux, M.; Kerrest, R. and Y. Montalanty (1973):** Etude de la surface foliare et de l'activite photosynthetique chez quelques hybrids de mais. *Ann. Amelior plantes*, 23: 95-107.
- [27] **Dubois, M ; Gilles, K.A ; Hamilton, J. K ; Rebens, P. A. and Smith, F. (1956):** Colorimetric methods for determination sugars and related substances. *Annals. Chem. Soc.*, 46:1662-1669.
- [28] **Duck, M.W; Gregg, B.M; Fernandez, R.T; Royal, D.H. and Cardoso, F.F. (2004):** Height control of *Picea spp.* and *Chamaecyparis lawsoniana* with uniconazole and 6-benzyladenine. *J. Enviro Hort.* 22, 165–169.
- [29] **El-Bassiony, A. M; Ghoname, A. A; El-Awadi, M. E; Fawzy, Z. F. and Gruda, N. (2012):** Ameliorative effects of brassinosteroids on growth and productivity of snap beans grown under high temperature. *Gesunde Pflanzen*, 64(4), 175-182.
- [30] **El-Emary, F.A. and Abd El-Aal, M. M. (2018):** Diverse responses for some growth biostimulants on some morphological, physiological, chemical, anatomical and yield characteristics of tomato plant (*Lycopersicon esculentum Mill*). *Archives of Agriculture Sciences Journal*, 1(2), 1-22.
- [31] **El-Emary, F. A; Abd El-Aal, M. M; Abd El-Dayem, H. M; Ismaeil, F. H. and Abou-Shlell, M. K. (2018):** Morphophysiological and anatomical studies on Moringa (*Moringa oliefera L.*). *Plant Archives*, 18(2): 1421-1442.
- [32] **El-Maadawy, E. I; Mansour, H. A. and Mostafa, H. G. (2006):** Response of annual flowering plants to manual pinching and benzyl adenine treatments: 1- Pot marigold (*Calendula affinalis L.*) plants. *J. Agric. Sci., Mansoura Univ.*, 31(7):4463-4483.
- [33] **El-Shafey, A.I; El-Feky, S.S. and Abo-Hamad, S.A. (2016):** Effect of sowing time and foliar application of yeast extract on growth and productivity of different cultivars of faba bean (*Vicia faba L.*). *Egypt J. Bo.*, 56: 35-48.
- [34] **El Sheikha, A. F; Allam, A. Y; Taha, M. and Varzakas, T. (2022):** How Does the Addition of Biostimulants Affect the Growth, Yield, and Quality Parameters of the Snap Bean (*Phaseolus vulgaris L.*)? How Is This Reflected in Its Nutritional Value?. *Applied Sciences*, 12(2), 776.
- [35] **El-Sherbeny, S. E; Khalil, M. Y and Hussein, M. S. (2007):** Growth and productivity of rue (*Ruta graveolens*) under different foliar fertilizers application. *Journal of Applied Sciences Research*, 3(5), 399-407.
- [36] **El-Tohamy, W.A. and El-Greadly, N.H.M. (2007):** Physiological responses, growth, yield and quality of snap beans in response to foliar application of yeast, vitamin E and zinc under sandy soil conditions. *Australian Journal of Basic and Applied Sciences*, 1(3): 294-299.
- [37] **Fadeel, A.A. (1962):** Location and properties of chloroplasts and pigment determination in roots. *Physiol. Plant.*, 15: 130-147.
- [38] **Faostat (2020):** Statistical database of the Food and Agriculture Organization of the United Nations, Rome, Italy. [http:// faostat.fao.org](http://faostat.fao.org).
- [39] **Fletcher, R. A. and Mccullagh, D. (1986):** A simple, sensitive, specific and steady bioassay for cytokinins. In *Bioassay and other special techniques for plant hormones and plant growth regulators*. Ed. Yapp, G.H.; L.H. Aung and G.L. Steffens. Published by Plant Growth Regulators Soc. of Amer., pp. 161-163.
- [40] **Furio, R. N; Salazar, S. M; Mariotti-Martínez, J. A; Martínez-Zamora, G. M; Coll, Y. and Díaz-Ricci, J. C. (2022):** Brassinosteroid Applications Enhance the Tolerance to Abiotic Stresses, Production and Quality of Strawberry Fruits. *Horticulturae*, 8(7): 572.
- [41] **Gabr, M. A; Fathi, M. A; Mohamed, A.I. and Mekhaeil, G. B. (2011):** Influences of some chemical substances used to induce early harvest of 'Canino' apricot trees. *Nature and Science*, 9(8), 59-65.
- [42] **Gad-Mervat, S. H. (2005):** Physiological studies on the effect of some growth regulators on soybean plant. M.Sc. Thesis, Fac. of Agric., of Moshtohor; Zagazig Univ., Egypt.
- [43] **Gao, S. (2020):** Function and Mechanism Study of Plant Cytokinins. In *Proceedings of the 2020 10<sup>th</sup> International Conference on Biomedical Engineering and Technology* (pp. 80-84).
- [44] **Gomes, M. M. A; Campostini, E; Leal, N. R; Viana, A. P; Ferraz, T. M; Siqueira, L. N; Rosa, R. C. C; Netto, A. T; Nunez Vazquez, M. and Zullo, M. A. T. (2006):** "Brassinosteroid analogue effect on the yield of yellow passion fruit plants", *Scientia Horticulturae*, Vol. 110, pp. 235–240.
- [45] **Hammad, S. A. R. (2008):** Physiological and anatomical studies on drought tolerance of pea plants by application of some natural extracts. *Ann. Agric. Sci., Ain Shams Univ., Cairo* 53 (2), 285–305.

- [46] **Haq, M. U. and Mallarino, A.P. (2005):** Responses of soybean grain oil and protein concentrations to foliar and soil fertilization. *Agron. J.*, 97: 910- 918
- [47] **Hayat, S; Ahmad, A. and Fariduddin, Q. (2003):** "Brassinosteroids: A Regulator of 21<sup>st</sup> Century". Hayat S, Ahmad A (eds.), *Brassinosteroids*, Springer, Netherlands, pp. 231–246.
- [48] **Holá, D; Rothová, O; Kočová, M; Kohout, L. and Kvasnica, M. (2010):** The effect of brassinosteroids on the morphology, development and yield of field-grown maize. *Plant Growth Regulation*, 61: 29–43.
- [49] **Horneck, D.A. and Hanson, D. (1998):** Determination of potassium and sodium by Flame Emission Spectrophotometry. In *Handbook of Reference Methods for Plant Analysis*, pp. 153-155.
- [50] **Horneck, D. A. and Miller, R. O. (1998):** Determination of total nitrogen in plant tissue. In *Handbook of Reference Methods for Plant Analysis*. Karla Y.P.(Ed.), pp. 75-83.
- [51] **Ibrahim, A.A; Magdy, A. S; Hassan, H. M. M. and Namich, A. A. M. (2016):** The influence of Lithovit fertilizer on the chemical constituents and yield characteristics of cotton plant under drought stress. *Int. J. Chem.Tech Res.* 9:1-11.
- [52] **Ikiba, L. K. (2017):** Effects of gibberellic acid and cytokinin application on morphological development, growth, quality and yield of french beans grown under different irrigation schedules (Doctoral dissertation, Doctoral dissertation, Kenyatta University, Kenya).
- [53] **Johanson, D. V. (1940):** *Plant microtechnique*. New York, London, McGrawHill Book Co. Inc. PP. 27-154
- [54] **Kamal, A. M. and Ghanem, K. M. (2012):** Impact of some bio-stimulants on organically cultivated snap bean plants. *Egypt. J. Appl. Sci.*, 27(2): 89-104.
- [55] **Kawamura, S; Tada, M. and Narsaki, T. (1966):** Sugar of cotyledon, hull and hypocotyls of soybean. *J. Jap. Soc. Food and Nutr.*, 19: 268.
- [56] **Khalil S. E. and Ismael, G. E. (2010):** Growth, yield and seed quality of lupinus termis as affected by different oil moisture levels and different ways of yeast application. *J. Am. Sci.*, 6, 141–153
- [57] **Khater, M. A; Zaki, F. S; Dawood, M. G; El-Din, K. G; Sadak, M. S; Shalaby, M. A. and El-Awadi, M. E. (2023):** Changes in some physiological and biochemical parameters of lupine plant via two bio-stimulents yeast extract and folic acid. *J. Mater. Environ. Sci.*, 14 (3), 373, 383.
- [58] **Khedr, Z. M. A. and Farid, S. (2000):** Response of naturally virus infected tomato plant to yeast extract and phosphoric acid application. *Annals of Agric. Sci. Moshtohor*, 38(2): 927-939.
- [59] **Koshioka, M; Harada, J; Noma, M; Sassa, T; Ogiama, K; Taylor, J. S; Rood, S. B; Legge, R. L. and Pharis, R. P. (1983):** Reversed phase C18 high performance liquid Chromatography of acidic and conjugated gibberellins. *J. Chromatogr.*, 256: 101-115.
- [60] **Liu, Y; Zhang, J; Guo, H; Zhao, A; Shao, D; Dong, Z. and Wang, P. (2020):** Effects of mulberry leaf and white kidney bean extract mix on postprandial glycaemic control in prediabetic subjects aged 45–65 years: A randomized controlled trial. *Journal of Functional Foods*, 73: 104-117.
- [61] **Mady, M. A. (2009):** Effect of foliar application with yeast extract and zinc on fruit setting and yield of faba bean (*Vicia faba* L). *J. Biol. Chem. Environ. Sci.*, 4(2): 109-127.
- [62] **Mahmoud, R. A; El-Desuki, M; Mona, M. A. M. and Aisha, H. A. (2013):** Effect of compost levels and yeast extract application on the pea plant growth, pod yield and quality. *J. Appl. Sci. Res.*, 9(1): 149-155.
- [63] **Mahmoud, H. I; Azzaz, N. A; Khalifa, Y. A; Mahmoud, M. A. and Fakhry, G. (2016):** Effect of foliar application with active yeast extract and benzyladenine on some vegetative growth criteria and chemical composition of lupine (*Lupinus termis* L.) plants. *Minia J Agric Res Develop*, 36: 193-214.
- [64] **Mahmoud, T.R. (2001):** Botanical studies on growth and germination of Magnolia (*Magnolia grandiflora* L.) Plants. M.Sc Thesis, Fac. Agric., Moshtohor, Zagazig Univ.,103pp
- [65] **Manea, A.I; AL-Bayati, H. J. and AL-Taey, D. K. A. (2019):** Impact of yeast extract, zinc sulphate and organic fertilizers spraying on potato growth and yield. *Res. on Crops*. 20 (1): 95-100. DOI : 10.31830/2348-
- [66] **Mohamed, A. S; Mohamed, M. H; Halawa, S. S and Saleh, S. A. (2023).** Partial exchange of mineral N fertilizer for common bean plants by organic N fertilizer in the presence of salicylic acid as foliar application. *Gesunde Pflanzen*, 1-12.
- [67] **Morsy, A. S. M; wadalla, A. A and Sherif, M. M. (2018):** Effect of irrigation, foliar spray with nano-fertilizer (lithovit) and N-levels on productivity and quality of durum wheat under Toshka conditions. *Assiut J. Agric. Sci.*, 49 (3): 1-26.
- [68] **Morsy, N. M; Abdel-Salam, M. A and Shams, A. S. (2018):** Comparing response of melon (*Cucumis melo*) to foliar spray of some different growth stimulants under two nitrogen fertilizer forms. *Egyptian Journal of Horticulture*, 45(1), 81-91.
- [69] **Muhammad, M. Q; Khursheed, M. Q. and Hasan, N. H. (2020):** Effect of foliar application with yeast extract and methanol on morphological



- and yield characteristics of faba bean (*Vicia Faba* L.). *Plant Arch*, 20(2): 9120-9124.
- [70] **Müller, B. and Sheen J. (2007):** Advances in cytokinin signaling. *Science* 318, 68–69.
- [71] **Müssig, C. (2005):** Brassionsteroid – promoted growth. *Plant Biol.*, 33 (2): 110 – 117.
- [72] **Nahed, G and Aziz, E (2007):** Stimulatory Effect of NPK Fertilizer and Benzyladenine on Growth and Chemical Constituents of *Codiaeum variegatum* L. *Plant. Am-Eurasian. J. Environ. Sci.*, 2(6): 711-719.
- [73] **Nassar, R. M; Ahmed, Y. M. and Nassar, D. M. (2011):** Effect of foliar spray with active yeast extract on morphological, anatomical and yield characteristics of kidney bean (*Phaseolus vulgaris* L.). *Australian Journal of Basic and Applied Sciences*, 5(5): 1071-1079.
- [74] **Nawar, D. A; Zyada, H. G. and Boghdady, M. S. (2018):** Effect Of Lithovit On Growth, Seed Yield Components And Anatomy Of Stem And Leaf Of Cowpea Plants Under Different Mineral Nitrogen Levels. *Zagazig Journal of Agricultural Research*, 45(3), 851-867.
- [75] **Neil, E. S; McGinley, J. N; Fitzgerald, V. K; Lauck, C. A; Tabke, J. A. and StreeterMcDonald, M. R. (2019):** White kidney bean (*Phaseolus vulgaris* L.) consumption reduces fat accumulation in a polygenic mouse model of obesity. *Nutrients*, 11(11): 2780.
- [76] **Nelson, N. (1944):** A protomorphic adaption of the Simonyi method for determine of glucose. *J. Biol. Chem.*, 153:373-377.
- [77] **Netwal, M; Choudhary, M. R; Jakhar, R; Devi, S. and Choudhary, S. (2018):** Exogenous application of Brassinoide and salicylic acid enhances on growth, yield and nutritional quality of Indian bean (*Lablab purpureus* L. var. *typicus*). *Journal of Pharmacognosy and Phytochemistry*, 7(6): 2093-2096.
- [78] **Nicander, B; Stahi, U; Bjorkman, P.O. and Tillberg, E. (1993):** Immunoaffinity co-purification of cytokinins and analysis by high-performance liquid chromatography with ultraviolet spectrum-detection. *Planta*, 189: 312- 320.
- [79] **Nolan, R; Shannon, O. M; Robinson, N; Joel, A; Houghton, D. and Malcomson, F. C. (2020):** It's No has bean: A review of the effects of white kidney bean extract on body composition and metabolic health. *Nutrients*, 12(5): 1398.
- [80] **Olaiya, C.O. (2010):** Presowing bioregulator seed treatments increase the seedling growth and yield of tomato (*Lycopersicon esculentum*). *J. Plant Growth Regul.*, 29, 349–356
- [81] **Osorno, J. M; Vander Wal, A. J; Posch, J; Simons, K; Grafton, K. F. and Pasche, J. S. (2020):** 'ND Whitetail', a new white kidney bean with high seed yield and intermediate resistance to white mold and bacterial blights. *Journal of Plant Registrations*, 14(2), 102–109.
- [82] **Piper, C.S. (1947):** Soil and plant analysis. The University of Adelaide, Australia.
- [83] **Punia, S; Dhull, S. B; Sandhu, K. S; Kaur, M. and Purewal, S. S. (2020):** Kidney bean (*Phaseolus vulgaris*) starch: A review. *Legume Science*, 2(3), e52.
- [84] **Qin, G; Wang, F; Liang, H; Tang, S; Shekh, K. and Wang, Y. (2019):** Subchronic study of a white kidney bean (*Phaseolus vulgaris*) extract with alpha-amylase inhibitory activity. *BioMed Research International*, 9272345.
- [85] **Reda, F. G. S. A; Baroty, G. S. A; Talaat, I. M; Abdel-Rahim, I. A. and Ayad, H. S. (2007):** Effect of some growth regulators and vitamins on essential oil, phenolic contents and activity of oxidoreductase enzymes of *Thymus vukgaris* L. *World Journal of Agricultural Sciences*, 3(5), 630-638.
- [86] **Rathna Priya, T. S. and Manickavasagan, A. (2020):** Common bean. *Pulses: Processing and Product Development*.:77-97.
- [87] **Raven, J. A. (2003):** Cycling silicon: the role of accumulation in plants. *New Phytologist*, 419-421.
- [88] **Saad, A; Abuzaid, A. S. and Rania, S. M. Eid. (2021):** Assessing Faba Bean Yield and Quality as Affected by Various Phosphorus Sources and Lithovit Levels. *Annals of Agricultural Science, Moshtohor*, 59(2), 817-830.
- [89] **Samuel, S. R; Deshmukh, P. S; Sairam, R. K. and Kushwaha, S. R. (2000):** Influence of benzyladenine application on yield and yield components in wheat genotypes under normal and late planting conditions. *Indian Journal of Plant Physiology*, 5(3), 240-243.
- [90] **Sandell, R. (1950):** Colorimetric determination of traces of metal. 2<sup>nd</sup> Ed. Interscience Publishers., Inc. New York.
- [91] **Sass, J. E. (1951):** Botanical Microtechnique. Iowa state college press, Ames, Iowa, pp. 228.
- [92] **Sasse, J. M. (2003):** Physiological actions of brassinosteroids: an update. *J. Plant. Gr. Regul.*, 22: 276-288
- [93] **Seadh, S. E; Attia, A. N. E; Badawi, M. A. and El-Hety, S. M. S. (2012):** Response of seed yield and its components of safflower to sowing dates, nitrogen fertilizer levels and times of foliar application with milagrow. *Journal of Biological Sciences*, 12(6), 342-348.
- [94] **Shallan, M.A; Hassan, H; Namich, A.A and Ibrahim, A. A. (2016):** The influence of lithovit fertilizer on the chemical constituents and yield characteristics of cotton plant under drought stress. *Int. J. of Chem. Tech. Res.*, 9 (8): 1-11.

- [95] **Shyaa, T. A. and Kisko, M. F. K. (2023):** Effect of Humic Acid, Cytokinin and Arginine on Growth and Yield Traits of Bean Plant *Phaseolus vulgaris* L. Under Salt Stress. *Baghdad Science Journal*.
- [96] **Snedecor, G. W. and W. G. Cochran (1980):** Statistical methods. 7<sup>th</sup> Ed. Iowa State Univ. Press Ames. Iowa, USA.
- [97] **Symons, G. M; Davies, C; Shavrukov, Y; Dry, I. B; Reid, J. B. and Thomas, M. R. (2006):** "Grapes in steroids. Brassionsteroid are involved in grape berry ripening", *Plant Physiology*, Vol. 140 No. 1, pp. 150–158.
- [98] **Taha, L. S; Ibrahim, S. M. and Aziz, N. G. A. (2016):** Vegetative growth, chemical composition, and flavonoids content of *Azadirachta indica* plants as affected by application of yeast natural extract. *Journal of Applied Pharmaceutical Science*, 6(4): 093-097.
- [99] **Taiz, L. and Zeiger, E. (2006):** *Plant Physiology*. 4<sup>th</sup> ed. Sinauer Associates Inc. Publishers, Massachusetts, US
- [100] **Tanaka, M; Thananunkul, D; Lee, T. C. and Chichester, C. O. (1975):** A simplified method for the quantitative determination of sucrose, raffinose and stachyose in legume seeds. *J. Food Sci.*, 40: 1087.
- [101] **Thorn, A. J. and Rogan, L. M. (2015):** The effects of lithovit on production and returns in a Hayward kiwifruit orchard. *Grower Services Manager*, 1:1-13
- [102] **Vogel, A. T. (1975):** A text book of practical organic chemistry. 3<sup>rd</sup> ed., Book society and Longmans group Ltd, London, 483-485.
- [103] **Wanas, A. L. (2002):** Resonance of faba bean (*Vicia faba* L.) plants to seed soaking application with natural yeast and carrot extracts. *Annals. Agric. Sci. Moshtohor*, 40 (1): 259-278.
- [104] **Wanas, A. L. (2007):** Effect Of Some Natural Extracts And Benzyladenine On Growth And Productivity Of Wheat (*Triticum Aestivum*, L.) Plants. *Journal of Plant Production*, Mans. Univ.,32(2): 1011-1029.
- [105] **Wang, C. F; Y.You; Chen, F. L. X. and Wang, J. (2004):** Adjusting effect of brassinolide and GA<sub>4</sub> on the orange growth. *Acta Agriculturae Jiangxiensis Universitatis*, 5-22.
- [106] **Wettstein, D. (1957):** Chlorophyll, Letal and dersubmicro-Spische Formmech Sell-derplastideu, *Exptl. Cell Res.*, 12, pp. 427
- [107] **Wichmann, G. and Basler, M. (2006):** Investigation of ingredients, product name LITHOVIT CO<sub>2</sub> foliar fertilizer for outdoor use. LEFO-Institut für Lebensmittel und Umweltforschung, untersuchung und Bewertung. Ahrensburg, Germany.
- [108] **Yang, Q. Q; Gan, R. Y; Zhang, D; Ge, Y. Y; Cheng, L. Z. and Corke, H. (2019):** Optimization of kidney bean antioxidants using RSM & ANN and characterization of antioxidant profile by UPLC-QTOF-MS. *Lebensmittel-Wissenschaft und -TechnologieFood Science and Technology*, 114, 108321
- [109] **Zakaria, M. A. T; Sakimin, S. Z; Juraimi, A. S; Ramlan, M. F; Jaafar, H. Z; Baghdadi, A. and Din, S. N. M. (2018):** Effect of different water regimes and plant growth regulators on growth, physiology and yield of banana (*Musa acuminata* cv. Berangan) in tropical climate. *Fundamental and Applied Agriculture*, 3(3): 505-514.
- [110] **Zewail, R. M. Y. (2014):** Effect of seaweed extract and amino acids on growth, productivity and some biocostituents of common bean (*Phaseolus vulgaris* l) plants. *J. Plant Production*, Mansoura Univ.,5 (8): 1441 – 1453.
- [111] **Zewail, R. M; Khedr, Z. M; Ismael, F. H. and Abd El hameed, E. F. (2019):** Enhancement of Soya bean (*Glycine max* L.) Plants growth, yield and seed quality by using putrescine, benzyladenine and yeast extract. *Annals of Agricultural Science*, Moshtohor, 57(2): 493-506.